

Evaluation of loxoprofen and its alcohol metabolites for potency and selectivity of inhibition of cyclooxygenase-2

Denis Riendeau,* Myriam Salem, Angela Styhler, Marc Ouellet,
Joseph A. Mancini and Chun Sing Li

Merck Frosst Centre for Therapeutic Research, 16711 Trans Canada Hwy, Kirkland, Quebec, Canada H9H 3L1

Received 20 October 2003; revised 11 December 2003; accepted 12 December 2003

Abstract—Loxoprofen, its *trans*-alcohol and *cis*-alcohol metabolites were evaluated for selectivity of inhibition of COX-2 over COX-1. The (2*S*,1'*R*,2'*S*)-*trans*-alcohol derivative was found to be the most active metabolite and to be a potent and nonselective inhibitor of COX-2 and COX-1 in both enzyme and human whole blood assays.

© 2004 Elsevier Ltd. All rights reserved.

Loxoprofen is a 2-arylpropionic acid anti-inflammatory agent with analgesic and anti-pyretic properties. It has been shown to be an effective inhibitor of the production of inflammatory prostaglandins *in vivo* but only a weak inhibitor of cyclooxygenase (COX) activity when tested using enzyme and cell assays *in vitro*.^{1–3} Its inhibitory effects on cyclooxygenase activity have been attributed to its (2*S*,1'*R*,2'*S*)-*trans*-alcohol reduction product (**5a**, see Scheme 1), the major metabolite of loxoprofen detected after administration.^{1–6}

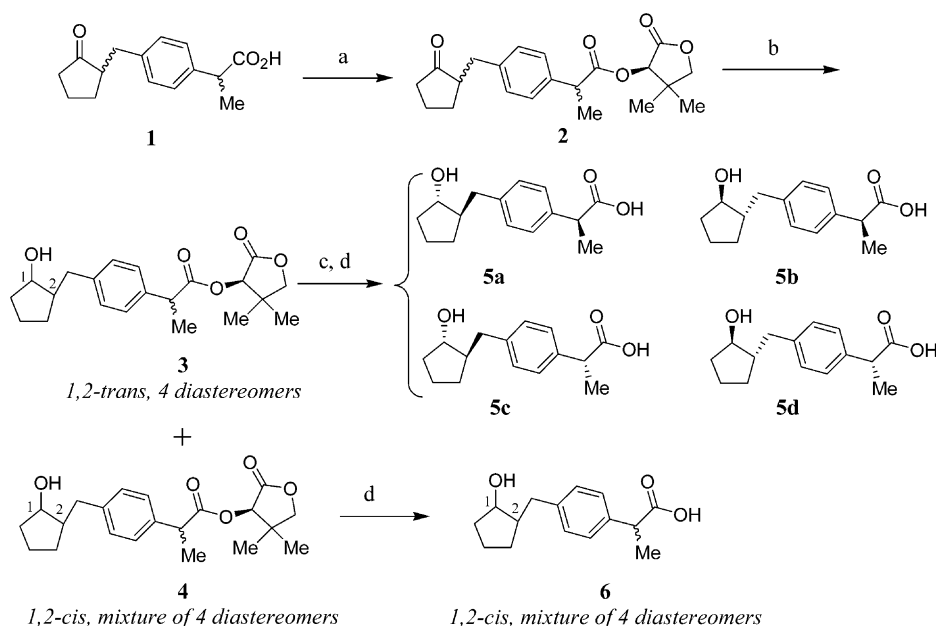
Assessing the selectivity of loxoprofen as a cyclooxygenase inhibitor is a difficult task since the compound is available as a mixture of 4 isomers, acts as pro-drug and has a rather complex profile of metabolites. Loxoprofen can be metabolized by stereoselective reduction, hydroxylation, chiral inversion, and conjugation to glucuronide derivatives.^{4–8} As described for several other prodrugs, the 2*R* propionic acid enantiomer of loxoprofen undergoes rapid chiral inversion to 2*S* after oral administration.^{7,8} A major route of metabolism for loxoprofen involves the stereoselective reduction of its cyclopentanone moiety to generate the (2*S*,1'*R*,2'*S*)-*trans*-alcohol derivative (**5a**). In addition to the *trans*-alcohol metabolite, reduction of the cyclopentanone can also yield the corresponding *cis*-alcohol (**6**), which has been detected mainly with the 2*S* configuration at the 2-aryl propionic chiral centre.^{7,8}

Previous studies have shown that the metabolite **5a** is more potent at inhibiting COX activity in ram seminal vesicle microsomes (a source of COX-1) than loxoprofen.¹ Limited data are available on selectivity. Inhibition of COX-2 mediated PGE₂ production by IL-1 stimulated human synovial cells (IC₅₀ = 0.12 μM) showed a 3-fold selectivity vs a COX-1 platelet assay for **5a**, about 10-fold higher than indomethacin and 10-fold lower than diclofenac.⁹ In the present study we have prepared the four *trans*-alcohols and a mixture of the *cis*-alcohol metabolites of loxoprofen and have evaluated them for their potency and selectivity of inhibition of human COX-2 over COX-1 using both enzyme and whole blood assays.

The four *trans*-cyclopentanol metabolites **5a–d** and the *cis*-cyclopentanol metabolite **6** were synthesized according to Scheme 1.¹⁰ Loxoprofen **1** was derivatized with (*R*)-pantolactone to give the ester intermediate **2**, which was reduced with sodium cyanoborohydride and separated by flash column chromatography to yield the *trans*-cyclopentanol **3** and the *cis*-cyclopentanol **4** in a ~5:1 ratio. The four diastereomers of **3** were then separated by chiral HPLC and the corresponding diastereomeric intermediates were subsequently hydrolyzed to provide the four *trans*-cyclopentanol metabolites **5a–d**.¹¹ The *cis*-cyclopentanol **4** was hydrolyzed in a similar manner to give the *cis*-cyclopentanol metabolite **6** as a mixture of four diastereomers.

The structures of the various metabolites are provided in Scheme 1 and were identified using the conventional

* Corresponding author. Tel.: +1-51-4428-2673; fax: +1-51-4428-4930; e-mail: denis_riendeau@merck.com



Scheme 1. (a) (*R*)-Pantolactone, 1-cyclohexyl-3-(2-morpholinoethyl)carbodiimide metho-*p*-toluenesulfate (CMC), DMAP; (b) NaBH₃CN, MeOH; (c) HPLC separation on Prep Chiral Pak AD column using 30% EtOH in hexane; (d) 0.4 M Na₂CO₃ in 50% aq MeOH, then 2 M HCl in HOAc; or 2 M HCl in 50% aq 1,4-dioxane.

names described by Takasaki and Tanaka.⁸ The IC₅₀ values for the inhibition of recombinant COX-1 and COX-2¹² by loxoprofen and its alcohol metabolites are summarized in the Table 1. Indomethacin and MF-tricyclic (a close analogue of rofecoxib¹³) were used as comparators for nonselective and selective COX-2 inhibitors. The (2*S*,1'*R*,2'*S*)-*trans*-alcohol derivative **5a** was found to be the most potent inhibitor and was equipotent at inhibiting the enzymatic activities of COX-2 (IC₅₀ = 0.39 μM) and of COX-1 (IC₅₀ = 0.5 μM) assayed under identical conditions. In contrast, the other *trans*-alcohols (**5a–d**), or loxoprofen itself, were much weaker or inactive as inhibitors of COX-2 with IC₅₀ > 100 μM. Similar data were obtained for COX-1, with a very weak inhibition detected for the (2*S*,1'*S*,2'*R*)-*trans*-alcohol (**5b**) (IC₅₀ = 51 μM). The *cis*-alcohol isomers (**6**) were not potent but gave an incomplete inhibition (~50%) of COX-1 and COX-2 at high doses (< 25 μM).

The selectivity of the (2*S*,1'*R*,2'*S*)-*trans*-alcohol derivative **5a**, the most potent metabolite, was further characterized using whole blood assays.¹⁴ In agreement with

the enzyme data, **5a** was a potent inhibitor of COX-2 (IC₅₀ = 0.3 μM) and showed no selectivity for the inhibition of COX-2 over COX-1 (IC₅₀ = 0.28 μM) (Table 2). As a comparison, the values for rofecoxib in these assay are IC₅₀ = 0.53 ± 0.02 μM for COX-2 and IC₅₀ = 18.8 ± 0.9 μM for COX-1.¹⁵

Loxoprofen and the other metabolites were also characterized for their effect on COX-1 or COX-2 in human whole blood assays. Although it is expected that the enantiomeric composition of the tested compounds might change during incubations *in vitro*,^{4,7,8,16} these assays were performed to detect potential inhibitory effects of loxoprofen and its metabolites in a more physiologically relevant milieu with cells using endogenously generated arachidonic acid substrate for the COX reaction. This assay would also detect alternative mechanisms of inhibition (such as inhibition of PLA₂). In the whole blood assay, loxoprofen was a very weak inhibitor of COX-2 (IC₅₀ of 13.5 ± 4.6 μM) and about 2-fold more potent against COX-1. Inhibitory effects on COX-2 and COX-1 were also detected with other metabolites (Table 2) but with a much lower potency than **5a**, possibly due to some isomerization to the most active **5a** metabolite during incubations with blood. The chirality of the 2-aryl propionic moiety is clearly important for the inhibitory effects on COX-2 or COX-1, (as reported for other 2-arylpropionyl acids¹⁶), with a large decrease in potency for the corresponding 2*R* enantiomers (**5c,d**). Considering their lower potency and the fact that they represent more minor metabolites than **5a** *in vivo*, the other metabolites would not be expected to contribute significantly to COX inhibition at normal effective doses.

In conclusion, the (2*S*,1'*R*,2'*S*)-*trans*-alcohol reduction product (**5a**) of loxoprofen is the most active major

Table 1. Inhibitory effects of loxoprofen and its alcohol metabolites on the activities of recombinant human COX-1 and COX-2 (mean ± SE)

	COX-2 IC ₅₀ (μM)	COX-1 IC ₅₀ (μM)
MF-tricyclic	0.23 ± 0.04	> 100
Indomethacin	0.73 ± 0.07	0.11 ± 0.02
Loxoprofen (1)	> 100	> 100
(2 <i>S</i> ,1' <i>R</i> ,2' <i>S</i>)- <i>trans</i> -alcohol (5a)	0.39 ± 0.05	0.5 ± 0.08
(2 <i>S</i> ,1' <i>S</i> ,2' <i>R</i>)- <i>trans</i> -alcohol (5b)	> 100 (30% Inh.)	51 ± 23
(2 <i>R</i> ,1' <i>R</i> ,2' <i>S</i>)- <i>trans</i> -alcohol (5c)	> 100	> 100
(2 <i>R</i> ,1' <i>S</i> ,2' <i>R</i>)- <i>trans</i> -alcohol (5d)	> 100	> 100
<i>cis</i> -alcohols (mixture of 4 isomers) (6)	> 25 (50% Inh.)	> 25 (44% Inh.)

Table 2. Potency of loxoprofen and its alcohol metabolites on COX-1 and COX-2 in human whole blood assays (mean±SE)

	COX-2 IC ₅₀ (μM)	COX-1 IC ₅₀ (μM)
MF-tricyclic	0.48±0.07 (n=16)	15.3±2.2 (n=22)
Loxoprofen (1)	13.5±4.6 (n=8)	6.5±1.3 (n=9)
(2S,1'R,2'S)- <i>trans</i> -alcohol (5a)	0.30±0.07 (n=7)	0.28±0.14 (n=5)
(2S,1'S,2'R)- <i>trans</i> -alcohol (5b)	2.0±1.3 (n=5)	4.7±1.4 (n=9)
(2R,1'R,2'S)- <i>trans</i> -alcohol (5c)	49% at 30 μM	20.2±4.3 (n=5)
(2R,1'S,2'R)- <i>trans</i> -alcohol (5d)	> 33	> 25
<i>cis</i> -alcohols (mixture of 4 isomers) (6)	22±10 (n=6)	1.9±0.3 (n=5)
Indomethacin ¹⁵	0.44±0.07	0.19±0.02
Rofecoxib ¹⁵	0.53±0.02	18.8±0.9

metabolite of loxoprofen with a potency of inhibition of COX-2 comparable to that of indomethacin or of potent selective COX-2 inhibitors. This result is consistent with the high efficacy of loxoprofen as an anti-inflammatory agent.^{1–3} However, **5a** showed no selectivity for the inhibition of COX-2 as compared to COX-1 as assessed in both enzyme and whole blood assays. Although the gastrointestinal tolerability of loxoprofen has been shown to be superior to indomethacin,^{3,17} the data on the nonselective inhibition are in agreement with the ability of loxoprofen to cause gastric lesions in animals at lower doses than selective COX-2 inhibitors.^{18,19}

Acknowledgements

We thank C. K. Lau (Merck Frosst) for the chiral HPLC separation of compound **3**.

References and notes

- Matsuda, K.; Tanaka, Y.; Ushiyama, S.; Ohnishi, K.; Yamazaki, M. *Biochem. Pharmacol.* **1984**, *33*, 2473.
- Sugimoto, M.; Kojima, T.; Asami, M.; Iizuka, Y.; Matsuda, K. *Biochem. Pharmacol.* **1991**, *42*, 2363.
- Terada, A.; Naruto, S.; Wachi, K.; Tanaka, S.; Iizuka, Y.; Misaka, E. *J. Med. Chem.* **1984**, *27*, 212.
- Naruto, S.; Tanaka, Y.; Hayashi, R.; Terada, A. *Chem. Pharm. Bull. (Tokyo)* **1984**, *32*, 258.
- Tanaka, Y.; Nishikawa, Y.; Hayashi, R. *Chem. Pharm. Bull. (Tokyo)* **1983**, *31*, 3656.
- Choo, K. S.; Kim, I. W.; Jung, J. K.; Suh, Y. G.; Chung, S. J.; Lee, M. H.; Shim, C. K. *J. Pharm. Biomed. Anal.* **2001**, *25*, 639.

- Nagashima, H.; Tanaka, Y.; Watanabe, H.; Hayashi, R.; Kawada, K. *Chem. Pharm. Bull. (Tokyo)* **1984**, *32*, 251.
- Takasaki, W.; Tanaka, Y. *Chirality* **1992**, *4*, 308.
- Kawai, S.; Nishida, S.; Kato, M.; Furumaya, Y.; Okamoto, R.; Koshino, T.; Mizushima, Y. *Eur. J. Pharmacol.* **1998**, *347*, 87.
- Synthesis of the eight possible metabolites of loxoprofen has been reported in: Naruto, S.; Terada, A. *Chem. Pharm. Bull.* **1983**, *31*, 4319.
- trans*-Cyclopentanol **5c** contaminated with ~5% of **5b** and *trans*-cyclopentanol **5d** contaminated with ~10% of **5c**.
- Enzyme assays were performed in 50 mM KPi pH 8.0, 1 μM heme, 1 mM phenol, with 10 μg/mL of COX-1 or COX-2 microsomal fractions. Test compounds were added as a 100-fold concentrated stock solution in DMSO to 100 μL buffer. After a 15 min preincubation, the reaction was initiated by the addition of 10 μL of 100 μM arachidonic acid. The enzyme reaction was allowed to proceed for 5 min at room temperature before being stopped by the addition of 10 μL 1 N HCl. PGE₂ levels were then determined by EIA. For details see: Percival, M. D.; Ouellet, M.; Vincent, C. J.; Yergey, J. A.; Kennedy, B. P.; O'Neill, G. P. *Arch. Biochem. Biophys.* **1994**, *315*, 111.
- Oshima, M.; Dinchuk, J. E.; Kargman, S. L.; Oshima, H.; Hancock, B.; Kwong, E.; Trzaskos, J. M.; Evans, J. F.; Taketo, M. M. *Cell* **1996**, *87*, 803.
- To study the inhibitory activities of these compounds on the two isoforms of cyclooxygenase (COX-1 and COX-2), human blood is either stimulated with lipopolysaccharide (LPS) for 24 h to induce COX-2 or the blood is allowed to clot spontaneously to activate COX-1. The production of prostaglandin E₂ (PGE₂) and thromboxane B₂ (TXB₂) are measured by immunoassay at the end of the incubation as readouts of COX-2 and COX-1 activity, respectively. For details see: Brideau, C.; Kargman, S.; Liu, S.; Dallob, A. L.; Ehrlich, E. W.; Rodger, I. W.; Chan, C. C. *Inflamm. Res.* **1996**, *45*, 68.
- Riendeau, D.; Percival, M. D.; Brideau, C.; Charleson, S.; Dube, D.; Ethier, D.; Falgoutyret, J. P.; Friesen, R. W.; Gordon, R.; Greig, G.; Guay, J.; Mancini, J.; Ouellet, M.; Wong, E.; Xu, L.; Boyce, S.; Visco, D.; Girard, Y.; Prasit, P.; Zamboni, R.; Rodger, I. W.; Gresser, M.; Ford-Hutchinson, A. W.; Young, R. N.; Chan, C. C. *J. Pharmacol. Exp. Ther.* **2001**, *296*, 558.
- Caldwell, J.; Hutt, A. J.; Fournel-Gigleux, S. *Biochem. Pharmacol.* **1988**, *37*, 105.
- Kawano, S.; Tsuji, S.; Hayashi, N.; Takei, Y.; Nagano, K.; Fusamoto, H.; Kamada, T. *J. Gastroenterol. Hepatol.* **1995**, *10*, 81.
- Arai, I.; Hamasaka, Y.; Futaki, N.; Takahashi, S.; Yoshikawa, K.; Higuchi, S.; Otomo, S. *Res. Commun. Chem. Pathol. Pharmacol.* **1993**, *81*, 259.
- Futaki, N.; Yoshikawa, K.; Hamasaka, Y.; Arai, I.; Higuchi, S.; Iizuka, H.; Otomo, S. *Gen. Pharmacol.* **1993**, *24*, 105.